



**BIOLOGICAL INDUSTRIES**  
**ISRAEL BEIT HAEMEK LTD.**

Kibbutz Beit Haemek 25115 Israel Tel. 972-(0)-4-9960595, Fax. 972-(0)-4-9968896, e-mail: info@bioind.com

## **Cell Proliferation Assay with XTT Reagent** **Cell Proliferation Kit**

**Cat. No.:** 20-300-1000 (for 1000 Assays)

**Storage:** -20°C

### **1. Introduction**

Cell proliferation assays are widely used in cell biology for the study of growth factors, cytokines and media components, for the screening of cytotoxic agents and for lymphocyte activation. The need for a reliable, sensitive and quantitative assay that would enable analysis of a large number of samples led to the development of methods, such as:

- use of radioactive thymidine to label DNA in live cells
- use of Brdu to label DNA in live cells (as a substitute for radioactive thymidine)

The above methods have a number of disadvantages, including: use of radioactive materials and relatively complex techniques. The use of tetrazolium salts, such as MTT, commenced in the 1950s and is based on the fact that live cells reduce tetrazolium salts into colored formazan compounds. The biochemical procedure is based on the activity of mitochondria enzymes which are inactivated shortly after cell death. This method was found to be very efficient in assessing the viability of cells. A colorimetric method based on the tetrazolium salt, XTT, was first described by P.A. Scudiero in 1988. Whilst the use of MTT produced a non-soluble formazan compound which necessitated dissolving the dye in order to measure it, the use of XTT produces a soluble dye. The use of XTT greatly simplifies the procedure of measuring proliferation, and is, therefore, an excellent solution to the quantitating of cells and their viability without using radioactive isotopes. This kit was developed to assay cell proliferation in reaction to different growth factors, cytokines and nutrient components. In addition, it is suitable for assaying cytotoxicity of materials such as TNF or other growth inhibitors. XTT can be used as a non-radioactive substitute for cytotoxic tests based on the release of  $^{51}\text{Cr}$  from cells with no less sensitivity.

The advantages of using this kit can be summarized with the following attributes:

- *easy-to-use*: there is no requirement for additional reagents and/or the cell washing procedures
- *speed*: multiwell plates and an ELISA reader can be used for reading
- *sensitivity*: can be assayed even in low cell concentrations
- *accuracy*: dye absorbance is proportional to the number of cells in each well
- *safety*: there is no need for radioactive isotopes

### **2. Kit Components**

#### **2.1 XTT Reagent (10x5ml)**

A sterile solution containing the XTT reagent. The solution should be stored frozen and should not be exposed to light. To avoid repeated re-freezing, dividing the solution into a number of vials after defrosting the original vial is recommended.

**Note:** if sediment is present in the solution, heat the solution to 37°C and swirl gently until a clear solution is obtained.

#### **2.2 Activation Reagent (2x0.5ml)**

A sterile solution containing PMS (N-methyl dibenzopyrazine methyl sulfate). The solution should be stored frozen and should not be exposed to light. To avoid repeated re-freezing, dividing the solution into a number of vials after defrosting the original vial is recommended.

**Note:** if sediment is present in the solution, heat the solution to 37°C and swirl gently until a clear solution is obtained.

### 3. Assay Principles

The assay is based on the ability of metabolic active cells to reduce the tetrazolium salt XTT to orange colored compounds of formazan. The dye formed is water soluble and the dye intensity can be read at a given wavelength with a spectrophotometer. The intensity of the dye is proportional to the number of metabolic active cells. The use of multiwell plates and an ELISA reader enables testing a large number of samples and obtaining easy and rapid results. The test procedure includes cultivation of cells in a 96-well plate, adding the XTT reagent and incubation for 2-24 hours. During incubation an orange color is formed, the intensity of which can be measured with a spectrophotometer, in this instance with an ELISA reader. The greater the number of active cells in the well, the greater the activity of mitochondria enzymes, and the higher the concentration of the dye formed, which can then be measured and quantitated.

### 4. Procedure

- 4.1 The cells should be cultivated in a flat 96-well plate. To each well add 100 $\mu$ l of growth media. The cells should be incubated in a CO<sub>2</sub> incubator at 37°C. In most cases cells can be used to assay proliferation after 24-96 hours.
- 4.2 Defrost the XTT reagent solution and the activation solution immediately prior to use in a 37°C bath. Swirl gently until clear solutions are obtained.
- 4.3 To prepare a reaction solution sufficient for one plate (96 wells), add 0.1ml activation solution to 5ml XTT reagent.
- 4.4 Add 50 $\mu$ l of the reaction solution to each well and incubate the plate in an incubator for 2-24 hours (usually, 2-5 hours are sufficient).
- 4.5 Shake the plate gently to evenly distribute the dye in the wells.
- 4.6 Measure the absorbance of the samples with a spectrophotometer (ELISA reader) at a wavelength of 450-500 nanometer. In order to measure reference absorbance (to measure non-specific readings), use a wavelength of 630-690 nanometer.

### 5. Notes

- 5.1 Defrost and prepare the reaction mixture only immediately prior to use.
- 5.2 Since the test is extremely sensitive, it is possible to use a low concentration of cells in the wells (approximately 5000 cells per well). Since there are cell types which show low metabolic activity, such as lymphocytes, kartinocytes and melanocytes, it is recommended to increase the concentration of cells to 2.5x10<sup>5</sup> cells per well, in order to obtain development of formazan color within a reasonable period of time.
- 5.3 Incubation time with the reaction mixture varies according to the type and concentration of the cells. Therefore, it is advisable to perform an initial test by reading the absorbance at various time lapses, i.e. after 4, 6, 8, 12 hours using the same plate.
- 5.4 Prior to reading the absorbance with a spectrophotometer, the plate should be gently shaken in order to evenly distribute the dye in the wells.
- 5.5 If the volume of the media in each well is larger than 100 $\mu$ l, add a larger amount of reaction mixture by the same increment (i.e. 100 $\mu$ l reaction mixture to 200 $\mu$ l growth media).

### 6. Summary of Proliferation Assay Using XTT Reagent: Flow Chart

- Defrost XTT reagent and activation reagent (37°C)
- Prepare reaction mixture (0.1ml activation reagent and 5ml XTT reagent for one plate)
- Add 50 $\mu$ l reaction mixture for each well (containing 100 $\mu$ l media)
- Incubate at 37°C for 2-24 hours (in most cases incubation for 2-5 hours is sufficient)
- Measure absorbance at a wavelength of 450-500 nanometer (reference absorbance: at a wavelength of 630-690 nanometer)

### 7. References

- \* Hansen, M.B., et al, (1989), *J. Immunol. Meth.* 119, 203-210
- \* Jost, L.M., et al, (1992), *J. Immunol. Meth.* 147, 153-165
- \* Roehm, N.W., et al, (1991), *J. Immunol. Meth.* 142, 257-265
- \* Scudiero, P.A., et al, (1988), *Cancer Res.* 48, 4827-4833
- \* Tada, H., et al, (1986), *J. Immunol. Meth.* 93, 157-165
- \* Weislow, O.S., et al, (1989), *J. Natl Cancer Inst.* 81, 577-586